TERMS AND CONDITIONS (INSTRUCTIONS) FOR VETERINARY PRACTITIONERS INVOLVED IN TESTING AND SAMPLING UNDER THE BOVINE TUBERCULOSIS ERADICATION AND BRUCELLOSIS MONITORING PROGRAMMES.

ACKNOWLEDGEMENT AND ACCEPTANCE OF THESE TERMS AND CONDITIONS (INSTRUCTIONS) IS REQUIRED FROM PRIVATE VETERINARY PRACTITIONERS (PVPs) AND WHOLETIME TEMPORARY VETERINARY INSPECTORS (WTVIs) AS THE APPLICATION FOR CONTINUED APPROVAL TO TEST/SAMPLE FOR THESE PROGRAMMES UNDER THE APPLICABLE LEGISLATION.

ACKNOWLEDGEMENT AND ACCEPTANCE OF THESE TERMS AND CONDITIONS (INSTRUCTIONS) SHOULD BE MADE ONLINE VIA THE ANIMAL HEALTH COMPUTER SYSTEM (PVP LINK)

FOR THOSE NOT OPERATING ONLINE THE ACKNOWLEDGEMENT/ACCEPTANCE (FORM ER4A) SHOULD BE SIGNED AND RETURNED TO THE REGIONAL VETERINARY OFFICE (RVO)

Please Note: If there is any element of this document which is unclear or which you do not understand please contact the SVI in your RVO to discuss the matter.
Index

1. Terms and Conditions attaching to Tuberculin testing and Brucellosis sampling .....3
1.1 Terms and Conditions applicable to Private Veterinary Practitioners (PVPs) for Eligibility to Test/Sample ..........................................................3
1.2 Specific Test Approval at herd/animal level .........................................................4
1.3 Requesting test approval at animal or herd level ..................................................4
1.4 A Herd Test ............................................................................................................6
1.5 Herd Profile ..........................................................................................................8
1.7 Administration of Veterinary Medicines in the context of Testing/Sampling ..........9
2. Number and volume of tests issued and completed ..............................................10
3. The Basis for Veterinary Certification and use of AHCS 1 amendment forms .... 10
3.1 Conflict of interest ..............................................................................................12
3.2 Newly qualified practitioners .............................................................................12
4. Handling Facilities and Farmer Assistance ............................................................13
5.1 Quality Control of Testing ...................................................................................13
5.2 Charge on PVPs for additional costs incurred by the Department .....................15
6.1 Payment of Fees ................................................................................................15
6.2 Tax Clearance Procedures ..................................................................................16
7. Data Protection .....................................................................................................16
8. Legislation .............................................................................................................17
9. Instructions for the Single Intradermal Comparative Tuberculin Test (SICTT) ...18
9.1 Purpose of the Test ..............................................................................................18
9.2 Intradermal Injection ..........................................................................................18
9.3 Equipment ..........................................................................................................19
9.4 Equipment checklist: ..........................................................................................21
9.5 Tuberculin ...........................................................................................................22
9.6 Detailed TB Test procedure ................................................................................22
9.6.1 Day 1 (Injection) ..............................................................................................22
9.6.2 Day 2 (72 hours +/- 4 hours post-injection)......................................................26
9.6.3 Testing for Export purposes ...........................................................................34
10. Instructions for Blood Sampling for Brucellosis ..................................................36
10.1 Animals to be tested .........................................................................................36
10.2 Equipment required ..........................................................................................36
10.3 Bleeding ............................................................................................................37
10.4 Packing of Samples .........................................................................................38
10.5 Description of herd and recording of data on animals sampled ......................39
10.6 Forwarding of Samples to Laboratory ..............................................................39
10.7 Supply of Blood Tubes .....................................................................................40
10.8 Disinfection ......................................................................................................40
10.9 Removal of Test Materials from Holding .........................................................40
10.10 Private Pre-Movement Testing .......................................................................40
10.11 General ............................................................................................................40
Appendix 1 To check handheld re untested animals ...............................................41
Appendix 2 Relevant Circulars ................................................................................42
Appendix 3 NOTES ON COMPLETION OF PRE PRINTED ER15B .......................46
Appendix 4 Cattle Passport ......................................................................................47
1. Terms and Conditions attaching to Tuberculin testing and Brucellosis sampling

1.1 Terms and Conditions applicable to Private Veterinary Practitioners (PVPs) for Eligibility to Test/Sample

i. For the purposes of these conditions/instructions “PVP” includes Wholetime Temporary Veterinary Inspectors.

ii. To conduct the single intradermal comparative tuberculin test (SICTT) and sampling for Brucellosis under the Animal Health and Welfare Act 2013, Regulations made thereunder and related EU legislation, PVPs are required to:

a) be entered in the current Register of Practitioners for Ireland,

b) be authorised by the Minister under the Act, and,

c) be approved under the TB Regulations and/or Brucellosis Regulations (Statutory Instruments considered to be Animal Health and Welfare Regulations) as appropriate.

d) commit formally and adhere strictly to the instructions, terms and conditions as laid down in this ER4 document; Approval to test takes place on an annual basis following acknowledgement/acceptance of the ER4 instructions either online or on receipt of the acknowledgement (Form ER4A) attached to the letter that accompanies the ER4 (off-line PVPs only). Approval/re-approval for testing is conditional on acknowledgment/acceptance of the ER4 instructions. Failure to acknowledge/accept ER4 terms and conditions will result in no approval or re-approval as the case may be.

e) use and update the unique identity codes (user code and password) and access number (Personal Identification number (PIN)) issued for personal use in respect of the Animal Health Computer System (AHCS) (where operating on-line) and keep details of these confidential. A PVP must not allow any third party access to their identity codes and/or PIN as this would facilitate false certification of a test.

iii. In addition to the above new applicants must:

a) apply to the RVO for approval to test by submitting form ER3. The RVO will then arrange a meeting with the SVI at which time the ER67 Contract will be signed by both the SVI and the new PVP;

b) attend a TB training course as prescribed by the Minister;

iv. No liability shall attach to the Minister for Agriculture, Food and the Marine for compensation or damages or costs in respect of any claims arising from the performance of testing/sampling under the Programmes.

v. Failure to comply with the instructions, terms and conditions, including those relating to equipment, performance of test, record keeping and other administrative procedures, may, depending on the nature of the infringement, result in sanction appropriate to the infringement up to and including the immediate withdrawal of approval to conduct the Single Intradermal Comparative Tuberculin Test (SICTT) and Blood sampling in accordance, as appropriate, with this ER4.
vi. The approval to test/sample may be terminated by notice of either party or following a
decision in the context of the Appeals Procedure referred to at a) below and subject to b)
below:

a) Disputes arising and regarding the performance of testing by an approved veterinary
practitioner (including withdrawal of authorisation to carry out tuberculin testing and
or blood sampling) shall be subject to the internal appeals procedures established by
the Department.

b) Notwithstanding the above, the decision of the Minister in relation to all aspects of
approval and termination thereof shall be final.

vii. A PVP who has been suspended from tuberculin/blood testing/sampling in Ireland may
likewise be suspended in Northern Ireland and vice versa. Acceptance of the ER4
conditions/instructions will be taken as consent to sharing information, with DARD
Northern Ireland.

1.2 Specific Test Approval at herd/animal level

i. Notwithstanding the authorisation and approval to test outlined above, tuberculin testing,
[the Single Intradermal Comparative Tuberculin Test (SICTT) for TB] or blood sampling
for TB or Brucellosis may not be conducted at animal (TB) or herd level (TB and
Brucellosis) without the Department's prior approval. An approved test is therefore a test
for which permission has been granted by the SVI or VI in charge at the RVO. Approval is
subject to the conditions specified, relating to issues such as date after which the test is to
be done, interpretation level, animals to be tested etc.

ii. Further details on the conditions specific for tuberculin testing and blood sampling are set
out below in sections 9 and 10 respectively.

iii. Fees will not be paid by the Department in respect of unapproved tests and such tests may
be invalidated by the Department.

iv. The PVP, having been issued with an approved test may, in exceptional circumstances,
request the SVI or the Veterinary Inspector (VI) in charge for permission not to carry out
the approved test issued.

v. If the PVP does not complete approved tests within the allotted time, or on the planned
dates submitted to the RVO, in the absence of a valid reason given in writing, and/or does
not submit test reports within the prescribed timeframes, further approved tests will not be
issued other than in exceptional circumstances.

1.3 Requesting test approval at animal or herd level

i. As stated in section 1.2, tests must have the prior approval of the Department and it is
illegal to carry out a test without such approval. Test approval, in every case, is subject to
the receipt of an advance itinerary, within the instructed timeframe.
ii. For TB, a SICTT may not be conducted at animal or herd level in advance of the date scheduled for the test, or other than in accordance with the conditions specified in the test listing. This is essential so that tests on herds, where status is being restored, comply with EU Directives to resume trading eligibility. Such tests should not be carried out prior to their scheduled date as this may necessitate further tests and result in prolonged restriction of the herd.

iii. Herd test itineraries may be submitted to the RVO through AHCS by midnight prior to the commencement of the test and up to mid-day on the Thursday of the week prior to commencement of the test in the case of non-electronic submissions.

iv. In very exceptional circumstances, substitutions to the advance itinerary, cancellations, alterations or change of PVP, are permitted even on the date of the test if notified to the RVO for approval prior to the commencement of the test. Communication by telephone is permitted in such very exceptional circumstances. PVPs must inform the RVO in cases where test reading has been arranged with the keeper to be carried out at a time other than that submitted on the advance itinerary using the e-mail facility; TBreadingtimechange@agriculture.gov.ie. Please note that this email facility relates only to change in reading time of test and not to change of planned date of test which should be done by resubmission of a new itinerary by midnight on the date prior to the date of test.

v. This e-mail address is not to be used for any other purpose. The only information to be input is as follows:

1. PVP Name
2. PVP Reference Code
3. Herd Number
4. Date of test reading
5. Time of Test reading as per itinerary
6. Proposed amended time of test reading
7. Reason for change in reading time

vi. Please ensure the details of change in time of reading include the above details as instructed. Failure to do so will appear as a non compliance on the PVP ER13A report.

vii. Sunday testing (commencement or completion) must, in particular, be notified to and receive individual approval from the SVI at the RVO prior to commencement.

viii. Itineraries for each private test with the planned date, time, and PVP must be submitted as part of the private test application for approval for online PVPs. Itineraries for private tests can be submitted on AHCS on or before the planned date prior to commencement of the test. Approval for each private test is only valid if the test is carried out in accordance with the itinerary submitted. Alterations to the submitted planned itinerary for private tests are facilitated by way of deletion of the original private test and submission of new private test approval application with the appropriate planned PVP, date and time. For PVPs working offline, test approval is subject to submission of form ER9 and receipt of approval from the RVO prior to commencement of the test. Permission for private tests may ordinarily, subject to disease exposure indicators, be granted for private tests to be carried out up to 30 days prior to the scheduled date of the Round test for herds that
completed a full herd test within the previous 12 months. The Round test may be also advanced if required by the RVO or requested by the keeper.

ix. All cancellations or alteration to the test itinerary must be forwarded to the RVO through AHCS (when interacting electronically) or by telephone and ER9 if still operating offline (not interacting electronically with AHCS). A current pre-printed or downloaded herd profile, obtained from AHCS (i.e. drawn down no earlier than one week prior to test commencement), must be used for each test. Download of the herd profile constitutes the approval for that test as per the conditions pertaining to the test listing and the advance itinerary submitted (appointed date and time).

x. It should be noted that any test, including private tests carried out other than as per the submitted advance itinerary i.e. at dates and times not corresponding to the planned date or by the planned PVP are not approved and therefore such tests are illegal. Such tests may not be accepted on the AHCS/AIM (the Animal Identification and Movement) systems and/or invalidated by the RVO therefore rendering test details invalid and thus preventing animal movement.

xi. It should be noted that non-compliance with the requirement to submit advance itineraries is a serious breach of instructions and may result in withdrawal of approval to test.

xii. Under no circumstance is tuberculin testing for TB (or blood sampling) permitted at or on a Mart premises. In the case of a herd-level test all animals on the holding are to be tested as part of the same herd (see section 1.4 and 1.5 below). PVPs, who are found to be in breach of this ruling, knowingly not testing all animals or testing animals under more than one herdnumber at the same location, will be sanctioned up to and including removal from the list of authorised Practitioners.

1.4 A Herd Test

i. The herd is all the bovine animals on the holding comprising an epidemiological unit. Whenever a full-herd SICTT is being conducted, the TB status of the herd will be updated when results are uploaded to AHCS and certified. The presence of bovine animals on the holding, known or apparent to the PVP, that are not presented for testing must be reported to the R.V.O. In the case of a herd test, when all eligible bovine animals on the holding are not tested, then de-facto the herd status may not be certified. In the case of blood sampling for Brucellosis, only those required by the Department to be sampled must be presented.

ii. Directive 64/432/EEC, as amended, requires that, when conducting a herd test in order to establish, retain or restore officially TB free status, all animals on the holding, with the exception of calves under six-weeks old which were born in the holding, must be subjected to routine tuberculin testing.

iii. Farmers are reminded in the test notification letter sent with each herd test scheduled that:

   a) All animals on the holding/in the herd must be presented at time of test,

   b) It is an offence to move animals out of the herd between the commencement and completion of the test,
c) Animals moved into the herd between commencement and completion of a full herd level test will require testing before the status of the herd (i.e. the status of all animals in the herd) may be certified.

iv. PVPs are provided with access to a copy of the material sent to their clients so that they may be aware of these requirements and that farmers have been so informed. Therefore, during the course of a full herd-test all animals, with the exception of calves under 6-weeks old that were born in the holding, must be tested, regardless of ownership, date of previous test or future plans to move, sell or slaughter them. The Department reserves the right to request presentation of any animal, regardless of age, for a test where deemed appropriate on veterinary grounds.

v. Please be advised that it is an offence on the part of the keeper not to present all animals on the holding for a herd test. Bovine animals kept on the same holding from part of the same epidemiological unit and therefore cannot be tested and certified under different herdnumbers. All sheds within the same yard are and have always been regarded as being on the same holding and therefore under a single herdnumber. See the “Information Note for Applicants for a herd/flock number for Cattle, Sheep and/or Goats” on the DAFM website which clearly states that it is not allowed to have animals under different herdnumbers in the same holding or the same yard even if in different fields on the farm or different sheds in the yard. Furthermore, farmers when being issued with a herdnumber formally undertake to maintain the herd as a discrete epidemiological unit and not intermingle with or keep on the holding stock under a different herdnumber. Farmers found to be in breach of this condition, will have their eligibility for a herdnumber reassessed by the RVO and if detected at IACS inspections will, additionally, ordinarily have a penalty applied to their BPS.

vi. Where a test is conducted in parts, all parts must be completed within 14-days of commencement of the first part to be considered a full herd test. Where a herd is being tested in parts with each part being commenced before the reading day of the previous part, the subsequent part must be created when the day 1 component of part 1 is uploaded on AHCS (after injection). Part tests being commenced after the reading day of the previous part can be created from the veterinary certification screen when the previous part is being signed-off. The Profile for the remaining untested animals is created automatically on AHCS when each part test is created and should be downloaded prior to commencing each new part test.

vii. Department pay tests will not be eligible for payment if not completed within the 14-days of commencement except with prior approval from the RVO. The test must be completed on all parts of the herd before the herd status can be certified and in particular before the individual animal test date can be inserted on the passport/identity card and returned to the keeper. The location of each part of the herd must be recorded and reported. Where the RVO has requested completion of the ER11 (Declaration by keeper of presentation of all animals), it must be presented to the keeper for signature and it is a legal offence if the keeper refuses to sign same.

viii. The same practitioner must conduct all parts of the test so as to be in a position to certify the status of the herd/animals. Only the RVO may sanction a deviation from this requirement.
ix. Once a herd test has commenced, it is an offence to move any animals off the holding to any other location (even for slaughter) until the test is complete and any animal on the holding intended for slaughter in the immediate future or animals tested in the recent past, or owned by others must also be tested for the herd test to be certified as completed.

1.5 Herd Profile

i. The herd profile provided from AHCS either pre-printed or downloaded electronically to the testing PVP by the Department is a list of all the animals present on the holding as notified to the Department by the keeper or his/her agent in compliance with the legal obligation to so notify.

ii. The PVP must have a copy (either electronic or hard copy) of the current herd profile whilst carrying out a herd test.

iii. The keeper must account for animals listed on the herd profile which are not presented for testing.

iv. It is possible to download the herd profile independent of the scheduling of a herd test. This may be to provide a keeper with a herd profile so that he/she may ensure any necessary updates/corrections to AIM are made and reflected in the herd profile or when permission has been given for a private test and the PVP wishes to use the herd profile to ensure accurate recording of tag numbers etc.

v. In order to safeguard the integrity of veterinary certification of the health status of the herd, the PVP must be satisfied that all animals in the herd have been presented for test.

vi. When conducting a full herd-test, if there are animals on the profile that have not been presented for test, the PVP must query the keeper as to the location of such animals and/or the reason for non-presentation for test (See section 9.6.2.6 below and Appendix 1). The reason given by the keeper must be duly recorded by the PVP and reported to the RVO via the test report (for information purposes only – i.e. not part of test certification).

vii. If other animals, not on the profile, are evident on the holding, as previously stated, these must be tested. When the certifying veterinary practitioner is satisfied on the basis of what is evident to him, including other animals visible or audible to the PVP in the same yard/sheds and/or adjacent fields/paddocks and otherwise the keeper’s information that all animals on the holding have been tested as required, then test certification may proceed.

viii. Where ‘missing’ animals are not adequately accounted for, the certifier (PVP) should present the keeper with an ER11 for completion (i.e. do not delay submitting the test report if the keeper cannot provide an answer immediately).

ix. Where animals, evident on the holding, are not presented for test and the keeper refuses to present them or claims that they are under a different herdnumber the PVP should contact the SVI/area VI as soon as practicable to notify of such refusal to present all animals. The SVI/VI will arrange for a VI to attend on day of test reading in order to ensure that the test for all animals is reported under one herdnumber.
x. When a test has been certified as complete and submitted to the Department, it shall be understood by veterinary officers of the Department that the veterinary practitioner is satisfied that all animals on the holding have been tested as required.

1.6 Testing facilities

i. Where a veterinary practitioner has concerns as to the facilities and assistance provided to enable him/her to conduct the test properly he/she should report such inadequacies to the RVO SVI/VI in charge of the area. The SVI/VI in the RVO shall act on such a report and maintain the confidentiality of the PVP as the person lodging the report. See also Section 6.1. Please also note the guidance provided by the Health and Safety Authority with respect to the obligation of Veterinary Practitioners and other professionals handling animals in farms, based on their professional training and experience, to ensure that safe systems of work are used. The HSA leaflet states “While the client or farmer should provide a safe place of work, there is a legal obligation on vets to ensure the work can be carried out safely. Vets must report any accident which causes their absence from work for more than three days to the Health and Safety Authority. Failure to report such accidents is an offence under the Safety, Health and Welfare at Work Act 2005. You can report an accident using form IR1 or online at www.hsa.ie. Where handling facilities or conditions are inadequate and potentially dangerous, the vet should raise their concerns with the farmer. If an adequate degree of safety cannot be provided the work should not continue until some remedy is put in place. Where vets have encountered circumstances where they have been at risk of serious injury and no remedy is foreseeable, they can report such a matter to the Health and Safety Authority. Details of the farm owner, location and any other specific information should be provided. If possible a photo would also be helpful. Confidentiality is assured: the source of the complaint will not be revealed and the matter can be dealt with in the course of a routine farm safety visit. You can contact the HSA on 1890 289 389 or email wcu@hsa.ie.”

ii. All surplus passports/identity cards should be submitted to the RVO with an accompanying ER124 (PVP Passport submission).

iii. Information supplied by the PVP on a test report may not be used to update AIM on behalf of the keeper. The reason given in the test report for absence of an animal on the profile serves only to confirm that the question was asked of the keeper and so facilitate test certification by the VI and it does not fulfil the keeper’s legal obligation to notify movements and deaths.

1.7 Administration of Veterinary Medicines in the context of Testing/Sampling

i. PVPs should not treat any cattle with a veterinary medicine in the course of carrying out the SICTT/sampling unless;

a) the medication is urgently required,

b) is unlikely to interfere with the cell-mediated response, and

c) the withdrawal period is likely to elapse before any reactor is required to be removed from the herd (unless there is no alternative treatment for the condition requiring urgent treatment).
ii. If the medication urgently required is to be administered by injection, the side of the neck and particularly areas adjacent to the sites used for the tuberculin test should be avoided. Furthermore, herd keepers should be advised by PVPs not to carry out routine treatment of animals immediately prior to or for the duration of a test. In cases where a prescription has been requested to coincide with a test, animals must not be treated until the individual animal test result is known. Such routine treatments will delay removal of reactor animals from the holding and prolong the restriction period accordingly. No compensation shall be payable, by the Department, for the additional testing or restriction period attributable to such treatments. Where routine treatments are necessary the test may, where it will not result in additional testing and/or delay in derestriction, be advanced with the permission from the RVO so that results are available before treatment.

iii. In line with best practice and for public health protection reasons if a PVP is aware that any reactors, which are intended for slaughter, have been treated with a veterinary medicine and that the withdrawal period will not have elapsed before the reactors are required to be removed from the holding, (s)he is requested to notify the RVO when reporting the reactors.

2. Number and volume of tests issued and completed

i. The number of approved tests issued to an approved PVP will be determined, at any particular time, by:
   a) the level of finances available to the Bovine TB Eradication Programme,
   b) farmer nominations, and,
   c) the requirements of the Programme.

ii. The volume of testing for TB performed by the PVP shall not, on a regular basis, exceed in any month, 750 animals per week, defined as six full-days testing amounting to approximately 250 animals injected or read per day. Note this is the number of animals regarded as the normal daily acceptable rate by the EU Commission throughout the EU.

3. The Basis for Veterinary Certification and use of AHCS 1 amendment forms

i. A certificate is a hand-written, printed or electronic statement of fact made with authority, whether or not it contains the word ‘certificate’. It is a grave offence if a veterinary practitioner issues any certificate which is untrue, misleading or improper.

ii. A certificate can only be issued after all the necessary steps have been taken to ascertain that the matters to be certified are in fact true.

iii. The legal bases for veterinary certification are the European Communities (Certification of Animals and Animal Products) Regulations 1999 (S.I. No. 380 of 1999), Regulation (EC) 882/2004, the TB Regulations and Brucellosis Regulations where relevant and, otherwise, the principles of certification as defined by the Veterinary Council apply.

iv. The veterinary practitioner carrying out a SICTT or blood sampling is required to ensure no conflict of interest exists or may be inferred (see 3.1 below).
v. The veterinary practitioner carrying out a SICTT or blood sampling is solely responsible for the accuracy of technique, recording, and is the only person who may certify any aspect of the test/sampling.

vi. The need for meticulous attention to detail cannot be too strongly emphasised. The injection of tuberculin in doses less than the prescribed amount (0.1ml) or at incorrect locations is likely to lead to infected cattle not being detected (bovine tuberculin) or conversely non-infected animals being deemed reactor (avian tuberculin).

vii. Meticulous testing/sampling technique and recording must, at all times, be clearly demonstrable as the basis for secure Veterinary Certification so as to ensure continued approval to test/sample. The veterinary practitioner’s certifying signature must be clearly legible on all reports (where such are not submitted electronically) or other certifying documents.

viii. In accordance with the principles of veterinary certification, Certification of a test shall be taken as confirmation by the veterinary practitioner that the test was performed in accordance with ER4 instructions, that, in the case of a herd test, the entire epidemiological unit was subjected to test in that as far as the certifier was aware, and based on the information supplied by the herdowner/keeper there were no other animals on the same holding that were not presented for test.

ix. Where tests are submitted electronically through AHCS, the veterinary practitioner who performed the test must personally sign off and certify the test on-line using the unique access codes and identity number assigned for that purpose. Certification is a reserved Veterinary function.

x. SICTT results and/or blood sampling details signed off and submitted to the Department shall be considered as certified by the veterinary practitioner and, as such, an accurate record of the facts as submitted.

xi. Subsequent requests for changes to original certification are regarded as a very serious matter and will only be considered when made by the original certifier and where appropriate justification, supported as necessary by documentary evidence, is provided. Form AHCS1 must be used to request a change where the test results are submitted electronically. The RVO will require explanation and/or evidence of the reason for the original error and/or details for validation of the change in certification. Animals will not be added to, deleted from, or have readings changed on a certified test unless there is documentary evidence to verify that the change in certification is fully supported and this has been accepted by Department veterinary staff.

xii. A PVP must not provide to a third party or allow others access to details of his/her unique identity codes and PIN issued for electronic certification purposes such that matters are purported to be certified by him/her. A PVP will at all times be responsible for the data entered, or edited under their unique code in the AHCS. In circumstances where this instruction has not been followed the PVP shall be held responsible for such false certification and liable for any consequences of such false certification. If a PVP believes that his/her AHCS password and/or PIN have become known to a third party s/he must immediately change the password and/or PIN. Administrative staff AHCS access codes provide access to all necessary areas excepting Veterinary certification.
xiii. A “herdnumber” is allocated to the “keeper” of a “herd” on a “holding” for the purposes of administering the TB Eradication Programme and the herdnumber so issued is also the registration number of the herd and the holding. The animals tested under a particular herdnumber may or may not be the property of the registered herdowner(s) and thus no inference as to ownership is conveyed by a test record or the presence of an animal on a particular herd profile. Under no circumstances should a test/sampling of animals be conducted under a herdnumber where it is clear that the animals are not located in that herd, on the holding occupied by that herd, at the time of test e.g. where animals are in a B&B holding, holding of a contract ‘rearer’, pound or elsewhere, they should not be reported as tested/sampled in the herdnumber or on the holding of the regular keeper/owner of the animals. The appropriate herdnumber in such cases is the herdnumber of the keeper in whose care and at which location the animals are currently held i.e. the herdnumber of the B&B holding, holding of a contract ‘rearer’, pound as relevant. In case of doubt, please contact the RVO to discuss the particular situation before returning to read the test or submit samples for analysis.

xiv. A VI in the RVO is ultimately responsible for test interpretation and assignation of herd status, and is therefore required to counter-certify the status of each herd/animal and various other matters pertaining to the test and to conformity with EU trading requirements etc. on the basis of the certification made by the testing PVP. A VI may avail of ancillary tests to support certification and decision making in this process.

3.1 Conflict of interest

i. The legislation cited at 3.iii prohibits certification of animals or products owned or from a holding, land or premises owned, in whole or in part by the certifier, their close relatives or by his veterinary partner/assistant. A close relative includes parents, parents in law, siblings, spouse/partner, and children and children of spouse/partner. The responsibility lies with the veterinary practitioner to ensure that no conflict of interest exists or may be inferred. Therefore, should a PVP become aware that he/she has been issued an approval to test in respect of which there is a conflict of interest under S.I. No. 380 of 1999, European Communities (Certification of Animals and Animal Products) Regulations 1999, or otherwise where a conflict of interest might be inferred, the onus is on the PVP to bring this fact to the notice of the SVI or the VI in charge. The PVP may not test the herd/animal(s) in question.

ii. See also the Veterinary Council of Ireland website www.vci.ie which states that Adherence to the Code of Professional Conduct is frequently more exacting than conformity with the law and also that veterinary practitioners should not issue a certificate which might raise questions of a possible conflict of interest.

3.2 Newly qualified practitioners

i. It should be noted that the Veterinary Council of Ireland, as part of the Code of Professional conduct, has drawn up official guidelines on the “Obligations of the employing Veterinary Practitioner” whereby Veterinary practitioners must provide appropriate professional support for newly qualified veterinary practitioners whom they employ. A newly qualified veterinary practitioner is one who is within the first year of graduation/registration. In addition, the Council also sets out “guidelines for new or returning graduates and their employers” which stresses the importance “that these
graduates/registrants are facilitated and supported so that they can develop further their day 1 graduation competencies in a structured progressive manner”. These guidelines are available on the Veterinary Council of Ireland website, www.vci.ie. The Department likewise expects a practice that employs a newly authorised graduate/registrant for the purpose of testing animals/herds for clients of the practice to arrange “technical and communication support” and supervision of the performance of the graduate “until both graduate and employer are satisfied they are competent and confident”. In the context of TB testing, a newly qualified veterinary practitioner is one who is within the first year of approval to conduct TB testing. The Principal/s of the practice should endeavour to ensure that testing carried out by employees of the practice can be conducted in compliance with ER4 annual instructions for testing.

4. Handling Facilities and Farmer Assistance

i. It is absolutely imperative that all animals to be tested are properly identified in accordance with legislative requirements and assembled in a yard, shed or paddock, located convenient to the testing facilities which must be such that the PVP can safely and effectively inject the tuberculin and conduct the reading and securely certify the result of the test. The PVP should not proceed with the test unless satisfied that the on-farm facilities are suitable and animals can be adequately restrained for the accurate performance of the test or sampling. As stated in Section 41 of the Animal Health and Welfare Act 2013, farmers are required to provide “assistance to an authorised officer, or person who accompanies the officer, and information to an authorised officer on request being made in that behalf by the officer, as the officer may reasonably require for the exercise of his or her functions under the Act” to ensure that the test/sampling can be performed accurately. In this context an authorised officer also includes a PVP approved to conduct TB/Br test/sampling. Assistance in restraining cattle is, ordinarily, essential to proper testing/sampling. The keeper should already have all cattle properly identified by means of a pair of official plastic ear tags (one in each ear). Testing of unidentified animals shall constitute grounds for withdrawal of testing approval.

5.1 Quality Control of Testing

i. DAFM, recognised as the Competent Authority by the EU, is obliged under EU legislation to ensure that tests are performed correctly for instance Article 3 of Directive 64/432/EEC requires that “Each Member State shall ensure that only animals that fulfil the relevant conditions laid down in this Directive are sent from its territory to that of another Member State” and Article 15 requires that “Member States shall take the appropriate specific measures to penalize any infringement of this Directive whether by a natural or a legal person.” In addition, Council Directive 96/93/EC on the certification of animals and animal products Article 5 states “(1) Member States shall introduce such checks and have such control measures taken as are necessary to prevent the issuing of false or misleading certification and the fraudulent production or use of certificates purported to be issued for the purposes of veterinary legislation (2) Without prejudice to any legal proceedings or penalties, the competent authorities shall carry out investigations or checks and take appropriate measures to penalize any instances of false or misleading certification which are brought to their attention. Such measures may include the temporary suspension of the certifying officers from their duties until the investigation is over.” In particular, if it is found in the course of the checks that: “a certifying officer has knowingly issued a fraudulent certificate, the competent authority shall take all necessary steps to ensure, as
far as is possible, that the person concerned cannot repeat the offence”. S.I. No. 380/1999 - European Communities (Certification of Animals and Animal Products) Regulations, 1999 transposes Council directive 96/93/EEC.

ii. As part of this Department’s routine quality control of the testing programme, test performance and results are monitored and assessed as a matter of routine. In addition, tuberculin storage and stock control will be monitored at practice premises inspections/visits.

iii. For each approved PVP conducting testing/sampling, the RVO will, amongst other things, arrange;

a) the supervision/inspection of equipment, test methodology and procedure including recording of data, biosecurity, cleansing and disinfection protocols while actually performing a test (ER13). Supervisions of test performance, will ordinarily be carried out unannounced at random, opportunistically or subject to risk based assessments, utilizing ER13A reports and other checks including those listed below, at a frequency as determined necessary by DAFM;

b) carrying out of check tests of herds;

c) carrying out of checks of animals after completion of a test;

d) monitoring recently tested animals for proper identification, the presence and location of SICTT clip marks, the presence, nature, location and accurate recording of reactions, sampling of animals nominated as tuberculin reactors for presence of a correlating Interferon-\(\gamma\) assay response and/or DNA match with blood sample submission, investigation of matters that come to the notice of the RVO post reporting of test (including where it becomes apparent that the same PVP has certified he/she has tested animals on the holding in the same yard.crush under separate herdnumbers); and

e) carrying out any other monitoring considered to be appropriate.

iv. In addition, when considering matters such as excessive numbers of animals tested in a day/week (noting the 250/day guideline in section 2.2), due cognisance shall be taken of other commitments that would reduce the number of animals considered acceptable e.g. regular meat inspection commitments, tuberculin usage, reactor disclosure rates, lesion disclosure rate in reactors and/or clear cattle, trace-back reactors in recently tested cattle, normal pattern of bovine and avian readings in reactor/non-reactor cattle, correlation with Interferon-\(\gamma\) assay results, insufficient/haemolysed/unsuitable submission rate, etc. will be subject to regular monitoring and comparison with the norm for the region.

v. Furthermore, the Department will conduct regular audits on administrative procedures such as test approval requests, advance itinerary submission and accuracy, compliance with legal requirements viz. a viz. accurate recording of identity details of animals/herds tested (taking into consideration animal movement records), prompt sample and/or test report submission, rate of submission of amendments to test reports, and other computer based procedures.

vi. A copy of the ER13A performance report, as generated from data routinely held on AHCS, will be made available to each PVP on a six monthly and annual basis and non
compliances identified therein will be followed up, as appropriate, including by unannounced test supervision, interview, warning, retraining, suspension from testing, prosecution and/or complaint to the Veterinary Council of Ireland. Details of non compliances and individual test performance are itemised on each ER13A report. Any PVP who is unclear as to the meaning or significance of matters on his/her ER13A or who wishes to clarify any matter thereon should contact the SVI in the RVO area to which the testing relates.

5.2 Charge on PVPs for additional costs incurred by the Department

i. Article 28 of Regulation EC Regulation 882/2004 is, by virtue of being a Regulation binding on a Member State. It requires “When the detection of non-compliance leads to official controls that exceed the competent authority's normal control activities, the competent authority shall charge the operators responsible for the non-compliance, or may charge the operator owning or keeping the goods at the time when the additional official controls are carried out, for the expenses arising from the additional official controls.” The FVO and European Court of Auditors, being auditors of compliance with EU legislation have confirmed that this Regulation covers the activities of PVPs engaged in testing under contract for the TB eradication programme. In light of this legal requirement, the Department is obliged to recover any costs incurred in cases where non compliance has necessitated additional follow up actions such as, testing, sampling, inspections or investigations as result of non-compliance by the PVP/practice with instructions set out in this document.

6.1 Payment of Fees

i. Fees calculated in accordance with the scale of fees applicable at that time shall be paid to the PVP who has conducted the test in respect of tests satisfactorily completed, including reporting timeframe, and which was nominated to be paid for by this Department. Tests performed outside the timeframe specified by the RVO and so notified to the farmer will not be paid for by the Department except where a case is accepted on a force majeure basis.

ii. Where a reactor is disclosed on a test nominated to be paid for by the farmer, DAFM will ordinarily assume responsibility for such payment subject to adherence with ER4 instructions. Exceptions include private tests and tests conducted on herds registered as feedlots where the herdowner has agreed with the Department to pay for one test annually and tests that have been declared null and void due to the detection of test irregularities.

iii. Where a PVP refuses to or finds he/she cannot proceed with testing because of inadequate handling facilities and/or assistance and certifies this to the RVO, the Department will pay the appropriate visit fee to the PVP and follow-up with the keeper as necessary.

iv. The Minister reserves the right to refuse or reduce payment to the PVP in respect of testing which was not carried out and/or not reported in accordance with this document and any other conditions of testing set down in legislation.
6.2 Tax Clearance Procedures

i. Under the terms of Department of Finance Circular 43/2006 ‘Tax Clearance Procedures Public Sector Contracts’, suppliers of goods and services are required to furnish a current valid Tax Clearance Certificate (TCC) if payments to them in any 12 month period exceed €10,000 (inclusive of VAT). The Circular 43/2006 is available at www.finance.gov.ie and from 1 January 2016 Revenue has ceased to issue paper tax clearance certificates. The paper certificates are replaced by a new electronic Tax Clearance Certificate (eTC) processing system. Paper certificates currently issued will remain valid until the date of expiry noted on those certificates. Further information in relation to this new electronic Tax Clearance Certificate processing system is available at www.revenue.ie

ii. Accordingly, all payments in excess of €10,000 will be withheld until such a certificate is provided. Alternatively, you may supply to the Department the psn/tax reference number and tax clearance access number as they appear on the electronic tax clearance (eTC) to facilitate online verification of your tax status. Delays in processing payments for tax reasons can be avoided in circumstances where the veterinary practitioner provides evidence to the Department that he/she is engaging with the Revenue Commissioners with a view to regularising his/her tax affairs.

iii. PVPs may opt to have any outstanding fees offset against their tax liability if this is acceptable to the Revenue Commissioners.

iv. In cases where a PVP is an employee within a practice, he/she must advise the local Revenue office when a source of income such as Department paid TB testing work (other than PAYE income) commences. This can be done by completing form TR1 – Tax Registration form for Sole Traders, Trusts and Partnerships available at http://www.revenue.ie/en/tax/it/leaflets/it10.html. PVPs should apply, once registered, for an electronic Tax Clearance Certificate.

7. Data Protection

i. The Department of Agriculture Food and the Marine is subject to the provisions of the Data Protection Acts 1988 and 2003. Under Section 8 of the Data Protection Acts 1988 and 2003, the Department is required to have PVPs sign an undertaking agreeing to the disclosure of information. PVPs are reminded that they too have a data protection duty towards their clients which extends to the herd profile downloads from and test details uploaded to AHCS.

ii. In relation to AHCS information, the Department is the “data controller” under the Data Protection Acts. The PVP is required to process personal data under the terms and conditions of his/her contract to conduct the single intradermal comparative tuberculin test (SICTT) and sampling for Brucellosis under the Animal Health and Welfare Act 2013, Regulations made thereunder and related EU legislation.

iii. The PVP’s role is that of “data processor”. The Department must ensure that the data protection standards are maintained and this includes by the data processor. The standards include appropriate security of data and data protection safeguards where personal data is involved.
iv. The Data Protection Acts requires the PVP to process personal data only on the basis of the authorisation and instructions received from the Department. Data may not be retained or used by the PVP for his/her own purposes.

v. PVPs as data processors must comply with the obligations imposed by section 2(1) (d) of the Data Protection Acts i.e. ensure that appropriate steps are taken against the accidental destruction, damage or loss of data.

vi. PVPs must ensure that sufficient guarantees are provided in respect of technical security measures to the personal data to protect it from unauthorised access or disclosures.

vii. The Department of Agriculture, Food and the Marine wish to confirm that the Data Protection undertaking/agreement also covers the following issues:

a) A commitment to provide prompt and full assistance to enable the Department to comply with any subject access request.

b) An agreement to inform ERAD Division immediately where there is any data security breaches in the data processor’s practice.

c) A right to engage in an adequacy audit and/or compliance audit to check compliance with the commitments in the agreement/contract.

d) On termination or expiry of the contract for any reason, all personal data held by the PVP as the “data processor” should be either returned to the data controller or deleted entirely from the data processor’s systems and files.

e) The Data Controller is required to be registered with the Data Protection Commissioner. The PVP as the Data Processor must also register with the Data Processors for the duration of the contract. You as the data processor on behalf of the Department should register with the Data Protection Commission as a Data Processor.

f) By signing the acknowledgement/acceptance of the ER4 conditions/instructions, it will also be understood that you as a data processor are committed to complying with the data protection rules set out in the Data Protection Acts 1988 – 2003 and the terms of the Departments Code of Practice.

g) Failure to comply with the provisions of the Data Protection Acts is an offence.

h) The details, including underlying reasons, of any sanction imposed by the Department in relation to TB and Brucellosis testing/sampling may be made available to the Department of Agriculture and Rural Development Northern Ireland for the purpose of ensuring that the Disease testing standards are upheld.

8. Legislation

1. Directive 64/432/EEC (as amended) on intra-community trade in bovine animals and swine.

2. Dir. 77/391/EEC introducing Community measures for the eradication of brucellosis, tuberculosis and leucosis in cattle.

3. Dir. 78/52/EEC establishing the Community criteria for national plans for the accelerated eradication of brucellosis, tuberculosis and enzootic leucosis in cattle.


9. Instructions for the Single Intradermal Comparative Tuberculin Test (SICTT)

i. DAFM has provided each authorised PVP with a copy of the instructional DVD “The Tuberculin Test” Copies are available for free on request from each RVO and ERAD HQ. The purpose of this recording is to demonstrate the correct application of the SICTT in accordance with EU Directive 64/432 and the requirements under the Irish Bovine TB Eradication Programme. PVPs who, when acknowledging reading/agreeing to comply with the ER4, acknowledge viewing of the DVD on conducting a test for TB, will receive CPD credit.

9.1 Purpose of the Test

i. to identify those cattle that are affected with bovine tuberculosis or capable of infecting other animals with bovine tuberculosis and to distinguish such animals from those that are not infected but which have become sensitised to bovine tuberculin as a result of exposure to cross-reactive antigens and

ii. in the case of a herd test, to determine the TB status of the herd in accordance with Directive 64/432/EEC and OIE requirements.

9.2 Intradermal Injection

i. The test, when carried out correctly, is highly reliable and has been assessed under Irish conditions as 90-98% sensitive and 99.95% specific.

ii. This reliability, however, is dependent upon the proper intradermal injection of both tuberculins (Bovine/Avian PPD) together with recording and reporting the accurate clinical observations on day 1 and with the accurate characterisation, measurement, and comparison of the reactions 72 hours later. Test reliability is also influenced by the volume of tuberculin administered and by the site of delivery of the tuberculin (both injections
should be in the same plane in the middle third of the neck on a line parallel to the blade of the scapula – see section 9.6.1.2 below). The site of delivery of tuberculin PPD is critical to the accuracy of the test. The neck was chosen, as the optimal intradermal injection site for sensitivity/specificity reasons1,2,3.

iii. The subcutaneous injection of tuberculin must be avoided as this will give rise to a false negative result in an infected animal and is also likely to lead to desensitisation of the site for a variable period.

iv. If the correct site on the side of the neck presented for test is unsuitable for some reason (scarring, old injection reactions etc.) tuberculin injection should be performed on the other side of the neck or if there is insufficient space for both injections in a suitable location on the same side of the neck one injection may be given on each side of the neck and a notation providing appropriate details made to that effect in the field book (See section 9.6.1.2).

9.3 Equipment

i. For quality control purposes all equipment must be presented when requested and surrendered for inspection/examination.

9.3.1 Syringes, Needles, Holsters

i. The syringes and needles used in the SICTT must be reserved for this purpose alone. Syringes must be clearly marked to distinguish between those used for avian tuberculin (red) and those used for bovine tuberculin (blue).

ii. Veterinary practitioners must have a minimum of 3, properly identified, working syringes in their possession and immediately available each day when planning to inject tuberculin for the purpose of conducting a SICTT. Spare needles, adaptors and identification thumb knobs must be carried at all times. Syringes must be emptied before commencing a test in a new herd. Syringes should be emptied at the end of each working day in order to prevent crystallisation of the tuberculin in the barrel, which could lead to syringe malfunction and consequential tuberculin administration inaccuracy.

iii. In order to qualify for testing under the programme each PVP must record on AHCS or manually submit certificates (if operating off-line) of purchase or service of at least two syringes within the previous 12-month period i.e. every calendar year each PVP must purchase or service at least two syringes.

iv. Syringes used for testing must be individually identified such that certification is specific to a particular syringe. Hence PVPs must ensure that any syringes in their possession at the performance of a SICTT conform to this requirement i.e. each syringe identified and either new or serviced no earlier than 30-months prior to the commencement date of the test being performed.

v. At all times syringes must be clean and in perfect working order.

vi. Before a test commences, it is essential to ensure that the loaded syringe is free of air and contains the correct tuberculin. The needle used should not protrude more than 3mm from the adaptor otherwise the tuberculin is likely to be injected subcutaneously. The needle should protrude at least 2 mm so that a successful intradermal injection is achieved.

vii. A fresh plug of cotton wool, soaked in methylated spirits, must be placed in each syringe holster at commencement of the tuberculin test in each herd such that the needle of the syringe will make contact with and rest in the methylated spirits between each injection.

9.3.2 Callipers, Clippers

i. Two callipers must be carried and maintained in good working order. Both lugs, together with the thumb-piece, must be stable and both the millimetre measurements and the reference mark must be clearly legible. A suitable clipping device, with a sharp cutting edge should be used and maintained in good working order. A suitable back-up device should also be carried.

9.3.3 Other Equipment

The following other items of equipment must be available at the time of the tuberculin test:

i. A field-book
   References will be made to the ‘field-book’ in this document and may be taken hereinafter as referring to any one of the following three approved recording methods:

   a) A hand-held computer operating a version of software approved by the Department with the current herd profile of the herd to be tested (i.e. drawn down no earlier than one week prior to test commencement) or,

   b) An official field book (ER14) and/or, the current herd profile of the herd to be tested (i.e. drawn down no earlier than one week prior to test commencement). When a hand-held computer is routinely used, in the event of a malfunction, a back-up manual recording system must be available when the test is being conducted i.e. Dept. supplied ER14 and wherewithal to write in it.

   c) All relevant details must be recorded contemporaneous to the conducting of the test and paper based records must be kept as the contemporaneous record for a period of not less than 6 years. Test records must be made available immediately upon request to a veterinary inspector or other authorised officer. PVPs are also advised to keep back-up records of electronic data (print-out, CD, DVD or other), being the contemporaneous record, for a period of 6 years. PVPs are advised that they may be required to attend court or other hearings and be required to produce such contemporaneous records and/or attest to details they have submitted to the Department.
ii. **Thermometer and stethoscope**
   A thermometer and stethoscope, appropriate for veterinary clinical examination of cattle.

iii. **Metal Ear Tags**
    Scheme metal ear tags (marked TT – temporary tag) are officially supplied by the Department for temporary ID purposes only and must be stored in a secure place until required. It is mandatory to have a supply immediately available together with a tagger (i.e. both physically present) when testing since performance of SICTT on un-identified animals is prohibited in law and will result in withdrawal of approval to test under the Disease Eradication Programme.

iv. **Reactor Tags/Discs**
    Reactor Tags, Red discs and Taggers for applying the tags described and as supplied by the Department.

v. **Protective Clothing/Disinfection**
    Boots and protective clothing and a supply of a disinfectant officially approved and effective against *M. bovis*. All personnel involved in testing are required to minimise the risk of the spread of infection, and to ensure that proper biosecurity and hygienic procedures, including disinfection, are carried out before entering and on leaving each farm/premises.

vi. All equipment should be clean and maintained in good working order.

### 9.4 Equipment checklist:
1. Boots, protective clothing and approved disinfectant;
2. Avian & Bovine tuberculin PPD;
3. Syringes x 3;
   a) 2 in use and 1 Spare syringe each certified no more than 30-months previously and provided certification has been submitted for 2 syringes, in the previous 12 months, in compliance with 9.3.1 above.
   b) identification thumb knobs,
   c) needles, adaptors and spanner to change needle;
4. Cotton wool & Methylated spirits;
5. Callipers x 2;
6. Clipping device;
7. Field book –(ER14) as supplied by the Department or the approved recording method for test;
8. ER 14 – a spare field book, as supplied by the Department, is required as back up even when electronic recording is the norm;
9. Thermometer & stethoscope;
10. Scheme metal ear tags and taggers – for identification of untagged animals – *mandatory for both days*;
11. Reactor Tags, taggers, ear-punch & red discs – as supplied by the Department for the identification of reactors.
12. All equipment should be clean and in good working order at commencement of test.
9.5 Tuberculin: – Bovine PPD and Avian PPD are supplied by the Department

i. Ensure that both tuberculins are within the expiry dates and record the batch number and expiry dates for each test (this may be relevant should there be a problem subsequently or legal challenge to the test). The tuberculin should be kept refrigerated between 2 to 8°C and protected from light until it is required for use. Not more than a single day’s supply of tuberculin should be kept un-refrigerated at any time. Tuberculin is provided free of charge and inefficient usage imposes an unnecessary cost on the State. Appropriate stock control procedures should be used to avoid product going out of date or other wastage.

ii. Tuberculin must only be used on the day on which the vial is opened. For biosecurity reasons, a vial partially used in one herd should not be used in another herd i.e. commence testing in each herd with an empty syringe and new vials. Used or partially used vials should be returned to the practice centre for safe disposal and should not be discarded on farms.

iii. The dose of each reagent is as follows: -
   - Avian tuberculin PPD: - 50 micrograms (0.1ml)
   - Bovine tuberculin PPD: - 100 micrograms (0.1ml)

9.5.1 Pharmacovigilance

i. As with all medicinal products an adverse reaction to tuberculin whether in a human or an animal must be reported to the Health Products Regulatory Authority (HPRS). Please copy such notifications to ERAD Headquarters.

9.6 Detailed TB Test procedure

9.6.1 Day 1 (Injection)

i. PVPs should remind keepers that no animals may be moved off the holding between day 1 and the reading day (even for slaughter unless certified in advance by the testing PVP as being an emergency welfare slaughter case or with the authority of the RVO). Untested/unread animals will lead to questions as to the appropriate status for the herd and the herd status will be suspended until appropriate tests can be performed. Similarly, the keeper will have been informed by the Department before the test is commenced that where a herd test is being performed, any animal that moves onto the holding between commencement and completion (reading) will require testing (part herd test) before the status of the herd (i.e. the status of all bovine animals on the holding) may be certified by the PVP. Where it comes to the notice of the testing PVP that such animals have been acquitted they should be tested as a further part-herd test.

ii. Before commencing a test the PVP must:
   a) Check that the facilities and assistance provided are as specified previously,
   b) Ensure that a fresh plug of cotton wool and methylated spirit has been placed in each holster, and,
c) Ensure that syringes are empty and syringes, needles etc. are free of any material that might constitute contamination from a previous herd.

9.6.1.1 Animal Identification

i. Before commencing a test (including reactor retests and tests where the herd is under restriction) the testing veterinary practitioner must request and take possession of all passports/identity cards in the keeper’s possession for the cattle on that holding.

ii. The PVP must be satisfied as to the identity of each animal being tested i.e. animal tagged and compatible with its description on the herd profile (breed, age, sex). The testing veterinary practitioner must also personally take the calliper readings for each animal on each day. When not personally recording the details of animal identity, test measurements etc., it is the responsibility of the testing veterinary practitioner, who will be expected to be in a position to certify the test, to assure him/herself of the accuracy of such recording. It is essential to record or verify each eartag number in full. Recording only the last 4-digits of animals not born in the herd is insufficient to ensure the accuracy necessary for test certification.

iii. Please check if details for animals not on the profile, which have been input as added animals correspond to the details on the animal’s passport (assuming the passport is recording the actual tagnumber in the animal’s ears). Any deviation from the details on the passport and/or database or changes made to the details on the profile e.g. sex, or breed will be considered as deliberate (certified by the testing veterinary practitioner) and an indication that the animal and passport/database records do not match. Please return excess (no animal presented), wrong passports (incorrect passport acquired with an animal) and passports that require amendment to the RVO. Where tests are submitted electronically, discrepancies in Gender, DOB, and Class will be highlighted prior to completion of certification.

iv. All animals on the herd-profile must be accounted for at a herd test. Animals left blank with no annotation, for information purposes, as to why they were not tested will raise discrepancy queries, at test interpretation, as to whether the full test is completed or not. Processing of incomplete tests will not be finalised by the veterinary inspector until such discrepancies are resolved. This will delay sign-off, test payment (where relevant) and eligibility to sell and export animals for the keeper.

v. Under no circumstances is the testing of unidentified animals permitted. When unidentified animals are presented for test they must, before testing or sampling, be tagged in the left ear, using temporary tags (brass and marked TT) as supplied by the Department and recorded on the test report. Testing or sampling of unidentified animals and certification of such tests is considered an extremely serious breach of testing and certification procedures, which may result in prosecution under the TB Regulations and/or Brucellosis Order, will result in the invalidation of the test in question and withdrawal of approval to test/sample and may be reported to the Veterinary Council. The presence of untagged/unregistered animals must be noted and procedures in Circular letters ER6A/2000 and ER16/2008 followed – copy attached at appendix 2.

vi. It is the keeper’s responsibility to correlate the temporary tagnumber inserted for test purposes with the tagnumber registered to the animal using the ER96 (Declaration in respect of bovines temporarily tagged). The keeper has been informed of this responsibility and that failure to have animals identified properly for test may result in test details, for the
animal(s) involved, not being certified by the testing veterinary practitioner until such time as they have satisfied themselves as to the correlation between the animal brass-tagged and the plastic tag number on the passport. This ordinarily will require re-visiting the herd to check the animal(s) involved. The herdowner/keeper will be responsible for any costs involved and (s)he has been so informed. The current ER96 has two sections, namely:

**Section A:** signed declaration by the keeper regarding the correlation of temporary brass tags with the permanent identity contained on plastic tags,

**Section B:** signed declaration by the testing veterinary practitioner that the correlation is correct and that he/she can certify the TB and/or Br test.

If Section A is completed, the RVO will consider the ‘discrepancy’ for the registered tagnumber resolved but unless Section B is completed and signed by the testing veterinary practitioner it will not record a test date on AHCS against that tagnumber. In addition, it will not return the passport to the keeper (assuming the passport has been submitted to it, as required, by the testing practitioner).

vii. Keepers should be strongly encouraged to order and replace missing plastic tags before the reading of the TB test is completed as this simplifies subsequent difficulties with certification and identity and obviates the need for completion of the ER96 form.

viii. The sooner keepers experiencing problems with the bovine identification regulations are identified the quicker and easier it is to resolve those problems and prevent the potential destruction of unidentifiable animals, development of animal welfare problems or Cross Compliance penalties. Thus, it is in the keeper’s best interest to ensure that the RVO is made aware of the difficulties at the earliest opportunity. If the PVP considers that a welfare problem may exist or could be anticipated the RVO should be notified and please consider if the Welfare ‘Early Warning System’ should be activated.

ix. At each location where animals or groups of animals are tested and the date(s) of part tests must be recorded and reported – this can provide considerable assistance in the event that an epidemiological investigation is required at any stage in the future. Clinical and other observations or other treatments likely to have a bearing on the results of the test must be recorded under Clinical Remarks on handheld devices or ER15B and, where appropriate, linked by tagnumber reference to the individual animal(s).

### 9.6.1.2 Site of Injection

i. As detailed in Section 9.2 above for accurate and consistent testing the injection site is critical. The approved injection sites specified by OIE and in Directive 64/432/EEC are situated at the border of the anterior and middle thirds of either side of the neck.

ii. The upper site (for avian tuberculin) shall be about four inches (10cm) from the crest of the neck.

iii. The lower site, for the injection of bovine tuberculin, should be about five inches (12.5cm) lower than the upper site, in the same plane on a line roughly parallel with the line of the shoulder/the ridge on the scapula (*the representational diagram below is a guideline only*).
iv. In calves under six weeks of age, or in young/small animals where there is insufficient space to separate the sites sufficiently on one side of the neck, one injection shall be made on each side of the neck (avian on the left, bovine on the right), at identical sites in the centre of the middle third of the neck.

v. For animals, which have non-associated scars/lesions/lumps/skin abnormalities or swellings adjacent to or obstructing the injection site(s) or leaving insufficient room to separate the sites on the presenting side of the neck the tuberculin should be injected into the opposite side and recorded in the ‘field book’ linked by tagnumber reference to the individual animal.

9.6.1.3 Site Preparation

i. It is required by OIE, EU and national legislation that the selected sites should be clipped (an area not less than 2.5cm in diameter) and cleansed (any dirt/debris removed), prior to injection. Specific injection site identification should thus be clearly apparent. The presence of any abnormalities at the injection site(s) should be recorded (not under Clinical remarks) in the ‘field book’ at the time and linked by tagnumber reference to the individual animal. The presence of skin tuberculosis should always be recorded as a clinical remark.

9.6.1.4 Measurement of Skin Thickness

i. Before injection, a fold of skin at each of the injection sites and within the clipped area shall be taken between the forefinger and thumb and accurately measured, using a callipers rounded up to the nearest millimetre; and the measurements recorded in the field book linked by tagnumber reference to the individual animal.

9.6.1.5 Injection Technique

i. A short sterile needle should be introduced, bevel edge outwards, into the skin in such a manner as to ensure the intradermal delivery of the tuberculin into the centre of the clipped site. This usually requires the insertion of the needle at a narrow angle to the skin. The insertion of the needle at a right angle to the skin will generally result in a subcutaneous injection being made. Such injections give rise to false negative results and must be avoided. Considerable pressure on the plunger of the syringe is usually necessary to make an intradermal injection. Absence of resistance to the flow of the tuberculin is an indication
that it has not been injected intradermally, or that the syringe is leaking or improperly loaded.

ii. Directive 64/432/EEC requires that “A correct injection shall be confirmed; by palpating a small pea-like swelling at each site of injection.” It does not allow a tolerance for non-palpation of a pea.

iii. If there is any doubt about either of the injections being delivered intradermally, a further injection should be made, preferably at a corresponding site on the other side of the neck. Such a procedure must be recorded in the ‘field book’ linked by tagnumber reference to the individual animal (in the Non clinical field and not under Clinical Remarks). At supervision the veterinary inspector shall pay particular attention to the successful production and palpation of the ‘pea’ delivered by a successful intradermal injection.

9.6.1.6 Recording of remarks:

i. **Tested animals:**
   On handheld devices, two fields exist for recording remarks against tested animals:
   
a) **Clinical Remarks (apparent to the VI at interpretation):**
   Entries under this heading, either in code abbreviation or text detail, should be confined to any clinical detail relating to an animal under test and which may have a bearing on the test result/interpretation and thus test certification (e.g. Skin TB, cough, snoring, gland enlargement, emaciation etc); any other detail i.e. reactor tag etc. should be recorded in the non-clinical field

b) **Non clinical field:**
   This should be used to record non-clinical details or reminders relating to the tested animal e.g. abnormality at injection site, reactor tag etc. or to serve as a reminder to the practitioner e.g. item for billing.

The ER15B only provides for clinical remarks; other remarks can be recorded on ER14.

ii. **Untested animals (including missing on day 2):**
   See above under animal identification. Any keeper information relating to animals on the herd profile, which is not presented for testing, should be recorded under **Clinical Remarks** (test certification issue). The lack of an annotation, for information purposes, against untested animals will raise discrepancy queries and will result in herd suspension (i.e. de-facto restriction) until RVO checks on missing animal(s) can be completed. Please see appendix 1 instructions re inspection of handheld for untested animals.

9.6.2 Day 2 (72 hours +/- 4 hours post-injection)

9.6.2.1 Reading of the Tuberculin Test

i. The SICTT must be completed by the same PVP who commenced the test, on the same holding, and using all the data recorded contemporaneously in the ‘field book’ on Day 1. Any departure from this must be for very exceptional reasons and must have the advance permission of the SVI/VI in charge in the RVO responsible for the issue of the herdnumber under which the SICTT is being conducted.
ii. Where, on test reading day, it becomes apparent to the testing PVP that there are other untested animals on the holding or the farmer (keeper) presents animals for test/reading under a different herdnumber on the same holding the PVP should contact the SVI/area VI at the first opportunity and should not falsely certify the test as if the entire epidemiological unit had been tested under the herdnumber on the test report. (See Sections 3 and 9.1 earlier).

iii. Each animal must again have its eartag number verified in full and its measurements, reactions, clinical symptoms/signs and any other observations immediately recorded in the ‘field book’ and correlated to the measurements and remarks recorded on Day 1 linked by tagnumber reference to the individual animal.

iv. Each site where tuberculin was injected must be examined, palpated, and measured. Measurements must be taken by first determining, by palpation, the nature of any response present and the broadest width of the response and then by carefully by placing the callipers across the broadest width of the response, without applying undue pressure, and recording the findings in the ‘field book’. All measurements must be rounded up to the next whole millimetre and recorded. Any additional remarks must be recorded immediately and correlated to the measurements and remarks recorded on Day 1 linked by tagnumber reference to the individual animal.

v. Clinical signs directly associated with all reactions to the tuberculin must also be recorded, contemporaneous with the time of reading, in the ‘field book’, linked by tagnumber reference to the individual animal. These signs include the presence of oedema, exudative necrosis, heat, pain or swelling at the individual injection site and/or heat, pain or swelling of the related prescapular lymph node.

vi. The presence of diffuse or extensive oedema, necrosis, heat, pain at the bovine injection site and/or swelling of the lymphatic ducts in the region or the related pre-scapular lymph node are regarded as clinical signs and always indicative of likely tuberculosis infection and the presence of such reactions must be reported. Animals showing such reactions to bovine tuberculin or with diffuse or extensive oedema, necrosis, heat or pain at the injection site must always be deemed as reactors, irrespective of the measurements recorded.

vii. The purpose in observing clinical signs, characterising reactions to the tuberculins and considering herd and animal histories and the status of contiguous herds is to facilitate the identification of animals, which may be infected but which have not been identified as reactors to the tuberculin i.e. False Negatives.

viii. The interpretation of reactions:

a) Positive: clinical signs or an increase of 4mm or more in skin-fold thickness.

b) Inconclusive: no clinical signs and the increase in skin-fold thickness is more than 2mm but less than 4mm.

c) Negative: if only limited swelling, with an increase of not more than 2 mm without clinical signs.
ix. Should the PVP suspect that the reaction is not a normal tuberculin response, he/she must contact a VI at the RVO at the earliest possible opportunity.

9.6.2.2 Interpretation of the test

i. Where 2 or more standard interpretation positives are found in a ‘clear’ herd (i.e. OTF status before the test) standard interpretation inconclusive reactors must be punched and tagged as reactors and recorded on the test report - unless instructed by the RVO to the contrary.

ii. Where the herd is undergoing a contiguous test, (high risk test because TB is present in a contiguous herd) or a 5a Special Check Test (5aSCT – a high risk test because TB in the herd in recent past) all standard interpretation inconclusive reactor animals must be identified for removal as reactors unless specific instructions in respect of the herd and interpretation have been received from the VI.

iii. Where the number of standard inconclusive reactors exceeds the number of standard reactors the RVO must be telephoned immediately and apprised of the situation. The RVO will then advise further punching and tagging of standard inconclusive animals as reactor.

a) **Standard Interpretation:**

Standard interpretation is applied when testing clear herds with a disease-free history or ‘restricted’ herds where an instruction has been received to apply standard interpretation. Standard interpretation of the single intradermal comparative tuberculin is as follows:

**Positive:** a positive bovine reaction that is more than 4mm greater than the avian reaction or the presence of clinical signs.

**Inconclusive:** a positive or inconclusive bovine reaction, which is from 1 to 4mm greater than the avian reaction and the absence of clinical signs.

**Negative:** a negative bovine reaction, or a positive or inconclusive reaction, which is equal to or less than a positive or inconclusive avian reaction and the absence of clinical signs in both cases.
b) **Severe Interpretation:**
Severe interpretation of the SICCT, to be used on specific instruction from the VI, or as above 9.6 1.2 (B) when 2 standard Reactors have been disclosed in a previously OTF herd, is as follows:

**Positive:** - a positive or inconclusive bovine reaction which is greater than the avian increase.

**Inconclusive:** - a positive or inconclusive bovine reaction, which is equal to, or 1 to 2mm less than, the avian reaction.

**Negative:** - a negative bovine reaction, or a positive or inconclusive bovine reaction, which is more than 2mm less than a positive or inconclusive avian reaction.

Animals displaying reactions to tuberculin, which cause them to be classified as reactors or inconclusive reactors must be identified and recorded as such in all cases.

---

**9.6.2.3 Identification of Reactors**

i. All animals classified as reactor must have a reactor tag, together with a red disc, inserted in the left ear for identification. The reactor tag and red disc serve as a visual warning as to the TB status of the animal for application of appropriate health and safety precautions, supply contract eligibility/controls and additional EU required post-mortem controls.

**9.6.2.4 Inconclusive Reactors**

i. When, as a result of the test, animal(s) are deemed as a standard interpretation inconclusive reactor(s) their respective passports/identity cards must be forwarded to the RVO and the keeper must be advised to isolate such inconclusive reactors from the herd pending re-test. Such inconclusive reactors should be kept separately from any reactors identified. The keeper must be informed by the testing veterinary surgeon that all movements out of the holding/herd are prohibited effective immediately – except for slaughter – until the RVO determines if the herd is eligible for a derogation to allow such movements within Ireland. Inconclusive reactor animals have EU required additional post-mortem controls and thus may only be slaughtered at approved premises – i.e. not local authority abattoirs.
ii. Inconclusive reactors should be classified in accordance with the above instructions and
the keeper advised immediately (see Advice to Keepers below).

iii. Where there is an inconclusive reactor animal in a herd, the herd disease status is restored
by way of the following:

   a) a clear test of the inconclusive reactor(s) after 42-days, (please note such an animal
      will remain confined to the herd of disclosure for its lifetime or until it is moved
directly for slaughter. In an exceptional case the RVO may permit one move to a
feedlot) or,

   b) slaughter, on permit at an approved premises, and laboratory examination of the
      inconclusive reactor(s) with negative results, or

   c) slaughter of the inconclusive reactor, on permit at an approved premises, followed
      by a clear herd test at a minimum of 42-days after removal of the inconclusive
      reactor(s).

iv. In certain cases, the RVO may allow the herd to trade on the domestic market under a
derogation provided for in Directive 64/432/EEC. Animals from such herds will be
precluded from export on the Animal Identification and Movement System (AIM) until the
clear disease status of the herd is restored as above or until the animal being exported has
been tested a minimum of 42-days following its departure from the herd in which there
was an inconclusive reactor.

v. As referred to in below in paragraph 9.6.3, where a standard inconclusive reactor animal is
present in a herd, under no circumstances (legal prohibition) will any animals in or from
the herd be approved for export until the clear status of the herd is confirmed by the RVO.

vi. Severe inconclusive reactors are marked as such. They will not ordinarily require a retest at
individual animal level or be removed if severe inconclusive again but, if considered
epidemiologically appropriate, a VI may remove them and/or any animal with a positive
bovine response as reactor e.g. if part of an infected group.

vii. The VI in the RVO will make the final interpretation of any test.

9.6.2.5 Animals Missing from Day 2

i. An animal, recorded in the ‘field-book’ as injected with tuberculin on Day 1 and for which
no Day 2 skin measurements or tuberculin response details is recorded, will be regarded as
not ‘read’ and the test will therefore be treated as incomplete and thus ordinarily herd
status may not be certified (as free). In such cases, the herd OTF status will, at a
minimum, be suspended until it can be clarified by means of test or otherwise. If an animal
recorded in the ‘field book’ on Day 1 is not presented for reading on Day 2, the keeper
should in the first instance be queried as to the absence of the animal, the explanation
recorded in the clinical remarks column and the RVO informed immediately (if the animal
died on farm the RVO may wish to arrange for examination of the injection site and/or a
post-mortem).
9.6.2.6 Advice to Keepers

When animal(s) are deemed reactor, including inconclusive reactor, the keepers must be advised:

i. to isolate them, pending slaughter on permit issued by the RVO (or retest in the case of an inconclusive reactor);

ii. that milk from a reactor/inconclusive reactor animals may not be used for any purpose (excepting feeding to a reactor) even if heat-treated and must not be supplied to the dairy/creamery;

iii. that milk from healthy animals belonging to reactor herds may not be used for the manufacture of heat-treated milk or for the manufacture of milk-based products unless it is first heat-treated at an establishment authorised by the Department;

iv. that the EU Requires keepers to retain a record of how much milk was produced by reactor and/or inconclusive reactor animals on the holding from the date of disclosure until the date of removal from the holding, how and where the milk was stored, how it was disposed of and the date it was disposed of;

v. that no reactor/inconclusive reactor on the holding may leave without permission from the RVO; and

vi. that non-reactor animals, in reactor herds, may be not be moved off the holding, except direct to slaughter on permit to an approved premises;

vii. that if the herd has an inconclusive reactor the RVO may allow trade within Ireland in which case a restriction notice will not be served but that (s)he should check with the RVO first if planning to move animals;

viii. that any animal disclosing a standard inconclusive reactor reaction will not be permitted to move from the holding concerned for the duration of its lifetime except to slaughter. RVOs may, if necessary, allow movement of inconclusive reactor animals that have passed the inconclusive reactor re-test, to a registered feedlot from where it shall go direct to slaughter within a reasonable timeframe;

ix. of their obligation to notify the PVP if any medicine has been administered to an animal that has been recorded as a reactor, the nature of the medicine and when it was administered.

9.6.2.7 Removal of Test Materials from Holdings

i. It is essential that the residue of all test materials employed in the test procedure including syringe parts, used tuberculin vials, needles, cartons and other items are gathered and removed at the time of leaving the holding.

ii. The safe and proper disposal of such materials in compliance with relevant legislation is the responsibility of the testing veterinary practitioner.
9.6.2.8 Completion of the ‘Field Book’, and Test report

i. Specific details on the completion of the three types of ‘field book’ are to be found at the front of the field book (ER 14) or from the appropriate software manuals for the handheld devices. Instructions for the completion of the pre-printed ER15B are attached at Appendix 3. If operating ‘online’ with AHCS, the test report must be completed, submitted and certified by uploading and interacting electronically with AHCS or if operating ‘off-line’ by completing submitting, certifying and returning the supplied pre-printed ER15b herd profile to the RVO.

9.6.2.9 Procedures relating to passports in the context of the disease eradication schemes:

i. When, an animal(s) is deemed as a standard interpretation inconclusive reactor(s) the passports/identity cards must be forwarded to the RVO pending retest of the inconclusive reactor or its slaughter on permit (EU laws for meat inspection mean that such animals may only be moved to slaughter on permit). See section 9.6.2.4 above.

ii. Passports for reactor animals or Inconclusive reactors must not be returned to or left with the keeper/herdowner (Instances where inconclusive reactors have been slaughtered at plants which have undertaken never to slaughter reactors has resulted in serious contractual consequences for the FBO).

iii. The passport/identity card for clear animals should be stamped with the date of the test by the testing veterinary practitioner except following a test where reactors are disclosed or a reactor retest that is not a clearance test. This date acts as an aide memoire to the farmer if planning to sell/move/slaughter the animal.

iv. The date of the test may be stamped under any of the three columns headed ‘Date of test’, ‘Herd No.’ or ‘Signature of Veterinary Surgeon’. See sample passport at Appendix 4 highlighting the columns that may be used.

v. Stamping the date of test on the passport/identity card is not deemed to be certification of the test. Certification of the test is completed by the testing veterinarian at the time the result of the test is submitted to the RVO by paper or electronically through AHCS.

vi. The only valid test date is that held on the Department’s Animal Health Computer System and it is this system that determines an animals’ eligibility for movement. It is thus extremely important that the most recent test is signed off on AHCS before movement of the animals.

vii. The Minister shall not be held liable in the event of disputes arising over incorrect test dates recorded on passports/identity cards.

viii. Veterinary practitioners are no longer required to submit passports/identity cards to the RVO except those relating to reactors (if requested), standard inconclusive reactors (see below and section D on page 23) and any surplus or incorrect passports/identity cards.

ix. In the event that any animal gives a reactor result to a test, the testing veterinary practitioner should retain custody of the passport/identity card for that animal for four months and unless its surrender has been requested by an authorised officer, to then destroy it. In addition, the testing veterinary practitioner must inform the keeper that the herd is restricted and that,
while the passports/identity cards for the remaining clear animals are being returned to him/her, (i) reactors including inconclusive reactors may only be moved directly to an approved slaughterhouse under a movement permit which must be surrendered to the slaughterhouse operative at time of presentation of the reactor and (ii) clear animals may be only moved directly to a slaughterhouse.

x. When the date of test has been inserted on the passports/identity cards for the clear animals, they should be returned to the keeper.

xi. Please note that keepers of herds restricted for TB (with limited exceptions) can move test negative animals direct to slaughter plants and abattoirs without the need for a movement permit as they will have the animal’s passport/identity card. However, keepers that knowingly move animals to non-allowed destinations (Marts and other farm holdings) will be subject to prosecution, loss of reactor compensation and cross compliance penalties.

9.6.2.10 Timely submission of Test Reports

i. The RVO should be informed on the day or at latest by the morning of the following working day, by telephone or otherwise, of all tests where reactors have been disclosed. Test reports (online/other as appropriate) must be submitted completed clearly and in full, including details of present/missing/surplus passports/identity cards. Surplus/other - passports/identity cards, as detailed in section 9.6.1.1 must also be returned at this time. However, excepting passports for reactors and inconclusive reactors, all other passports/identity cards should be stamped with date of test only, as previously described, and returned to the keeper.

ii. The test report on which a reactor or inconclusive reactor is disclosed must, by law, be submitted so as to reach the RVO not later than three (3) working days after the completion of the test.

iii. If any reactors have been medicated and have an unexpired withdrawal period this should be noted in the clinical remarks columns.

iv. Test reports on which no reactors are disclosed must, by law, be submitted to reach the RVO within seven (7) working days of test completion.

v. All test data and, therefore, export eligibility is relayed to marts and meat plants via the AHCS system. Failure to promptly upload and sign off on tests will result in rejection of animals at sales, export points or slaughter plants. It is essential that test reports for animals, which are to be moved or slaughtered, are submitted immediately. Animals on unreported tests will not have export status annotated/displayed at point of sale and/or may have a farm-to-farm compliance certificate refused should the test report not be submitted to the RVO and/or on AHCS before the animal is presented at the mart or the certificate requested.

vi. The Interpretation Act 2005 provides as follows:

“Periods of time: Where a period of time is expressed to begin on or be reckoned from a particular day, that day shall be deemed to be included in the period and, where a period of time is expressed to end on or be reckoned to a particular day, that day shall be deemed to be included in the period.” Thus both the test reading day and date of sign off on AHCS or receipt in RVO are each counted as days for the purpose of reckoning the submission interval and compliance with legislation.
vii. Details of temporary tags etc. must be recorded on the report, against the relevant existing animal on the profile if possible and all missing passports/identity cards recorded; surplus passports/identity cards must be submitted to the RVO within the timeframes detailed above. Where details for animals added to the profile do not correspond to the details on the animal’s passport, this will be considered as deliberate certification and an indication that the animal and passport (AIM) do not match and will consequently raise a discrepancy against the animal.

viii. Passports/identity cards for ‘clear’ animals should be stamped with the date of test after each clear test and returned to the keeper. Passports/identity cards should not be stamped for reactor retests except where it is a clearance test.

ix. Where a TB test is carried out in conjunction with a Brucellosis test, passports/identity cards for all Brucellosis eligible cattle must be retained until the result of the Brucellosis test is available and otherwise passports in respect of Brucellosis tests should be treated as outlined above in the same manner as following TB tests.

x. Be advised that the VI is ultimately legally responsible for test interpretation and the determination of animal and herd status. The RVO will notify the PVP in writing of any changes to the field interpretation of test results.

xi. The post mortem results of animals from herds tested by the PVP and which were slaughtered as reactors are notified on AHCS and available for access by the testing PVP.

9.6.2.11 No Stock Reports

i. The RVO will check the database prior to issuing herds for test. Herds with no stock on record will not be issued to a PVP for test until such time as the database indicates that stock have moved into the herd. In cases where a keeper who has been listed for a herd test and has been notified by you to test has no bovine animals, a no stock report should also be submitted by the due date on the listing to the RVO. This will avoid further queries from the RVO re untested herds. The RVO will then deal directly with the keeper as appropriate to regularise the position on the database.

9.6.3 Testing for Export purposes

i. An animal will only be eligible for export when it has passed the SICTT within the previous 30-days, there been no increase in skin-fold thickness of greater than 2mm present at the bovine site and there are no clinical signs such as diffuse or extensive oedema, exudation, necrosis, pain or inflammation of the lymphatic ducts in that region or of the lymph nodes (EU Directive 64/432). In all circumstances, other than a full herd-test, a tuberculin test may only be conducted if the interval to the previous tuberculin test (injection day to injection day) is a minimum of 42 days. In circumstances where an animal is tested at a herd test at an interval of less than 42 days from its previous test, the animal may be eligible for export if:

   a) the date of the initial test is within 30 days of the date of export
   b) the initial test result for the animal was such that it was export eligible (see first sentence above) and
c) provided the animal has not otherwise become ineligible by virtue of reactor or an
inconclusive reactor disclosure in the current herd or the animal itself exhibiting a
positive response to bovine tuberculin.

ii. Individual animal test data as reported and recorded on AHCS is automatically checked at
marts and export assembly centres to ensure that only animals which fulfil the export test
criteria as above are either indicated as eligible for or certified for intra-community trade.
It is therefore essential that tests are signed off on AHCS prior to presentation of animals at
marts and particularly for export. Animals are ineligible for export health certification if a
test is not certified/signed off by the PVP (reserved Veterinary function) on AHCS when
presented for export. Accordingly, to be eligible for export, permission for any necessary
pre-movement test must have been received, the test conducted in accordance with the
permission received and tests results uploaded by the PVP to AHCS or, for PVPs working
off-line, received in the relevant RVO at a minimum one full working day prior to the
intended export.

iii. Where a standard inconclusive reactor animal is present in a herd, under no circumstances
(legal prohibition) will any animals in or from the herd be approved for export until the
clear disease status of the herd is confirmed by the RVO.
10. **Instructions for Blood Sampling for Brucellosis.**

10.1 **Animals to be tested**

i. Following achievement of officially Brucellosis free status on the whole island of Ireland there is now no compulsory routine on farm testing for the disease. The disease will continue to be compulsorily notifiable and the Department will continue with appropriate monitoring measures for Brucellosis, such as testing culled cows at slaughter plants, aborted foetuses sent to Regional Veterinary Laboratories and post-abortion blood samples at no cost to farmers as part of its commitment to animal health surveillance in support of the livestock industry.

ii. The Department reserves the right to request any animal to be presented for test where deemed appropriate on veterinary grounds.

iii. The Brucellosis Order defines an "eligible animal" as any animal aged 12 months or more except a castrate. Following Ireland’s attainment of Official Brucellosis Free (OBF) Status in 2009, the policy from 2015 onwards is to confine testing to post-abortion tests, testing following indications of positive results at slaughter in animals culled from the herd and certain risk situations. Keepers of all herds requiring test will be notified in writing of the arrangements.

iv. The legal obligation to report all abortions in cattle remains unaltered as does the obligation to submit such fetuses, if available, for testing for Brucellosis to the Department’s Regional Veterinary Laboratory and/or to have the animal that aborted sampled for Brucellosis as soon as possible by a Veterinary Practitioner.

v. Under the Brucellosis monitoring programme some risk based testing will be scheduled in particular for female animals that may have been imported from non-OBF countries and will maintain a potential risk of latent infection until a post calving test clarifies their status. Such animals may be ‘flagged’ on the download from AHCS to alert you to sample them when presented for TB testing.

vi. However, the Department reserves the right to request that any animal be presented for test where deemed appropriate on veterinary grounds.

vii. Each animal presented at the test must be correctly identified according to its tag number, breed, sex, type, stage of pregnancy, age and abortion history. All these details must be recorded in the field book.

viii. Passports/identity cards for all Brucellosis eligible animals must be retained until the result of the Brucellosis test is available and otherwise passports in respect of Brucellosis tests should be treated in the same manner as following TB tests – see section H page 24. In the case of a post abortion test(s), the passport/identity card must be returned immediately to the RVO.

10.2 **Equipment required**

i. A small number of blood testing kits will be supplied by the Department to each practice to facilitate the limited testing described above. This kit is for sampling herds for Brucellosis testing only and is not to be used for sampling for other purposes. Replacement kits will
be provided based on the volumes of samples submitted under the Brucellosis monitoring programme. Surplus empty and unused sample tubes should not be submitted to the laboratory. Additional empty boxes can be ordered from the Blood Testing laboratory to enable you to use surplus empty sample tubes.

ii. To record relevant details of the test, you may use a hand-held computer (with Department approved software), an official field book (ER14) or a computer printout. All relevant details must be recorded at the time of the test and the record kept for a period of not less than 7 years. (For those who are operating AHCS on-line, an electronic record is automatically generated and stored). Paper based records and back-up records, being the contemporaneous records, should be kept for a period of 7 years. Even when a hand-held computer is routinely used, you must have an ER14 manual recording system as a back-up. The term ‘field-book’ refers to any of these approved recording methods.

iii. Tagging equipment must be clean and in good working order.

iv. Scheme metal ear tags (for temporary use for test ID purposes only) stored in a secure place until required (please refer to instructions issued in regard to the Farmer Plastic Tagging System).

v. Boots and protective clothing. To minimise the risk of the spread of infection, boots and protective clothing must be cleaned and disinfected on entering and leaving each farm.

vi. A supply of an officially approved disinfectant effective against brucellosis. A bucket and suitable hard brush should also be carried to facilitate cleaning and disinfection of boots.

10.3 Bleeding

i. A separate needle must be used for each animal to avoid cross contamination. Ensure that the tube fills to at least 3/4 of its capacity. Affix a pre-coded label to the upper 1/3 of the tube.

**Level of blood in blood tubes required for Brucellosis testing**

![Diagram showing level of blood in blood tubes required for Brucellosis testing]

**Place Erad Label Here**

**Min Blood Level**

7cm approx.
ii. It is essential to correctly correlate the sample identification number (tube code) to the animal identification in the field book. To minimise the possibility of error, this correlation should be carried out at the time of sampling. The person taking the blood samples is solely responsible for the accuracy of this correlation. The accuracy of sample identification and sample correlation must, at all times, be demonstrable as the basis for Veterinary Certification.

iii. No payment shall be made where insufficient blood is submitted and be advised that no visit fee will be payable to resample in such cases.

### 10.4 Packing of Samples

i. Blood samples tubes should be placed in the correct order in the aeroboard mould in sequential order from left to right in each row. (See diagram below). As above the person who took the blood samples is solely responsible for ensuring the accuracy of the identification of the samples and the correct correlation between the samples and the animal sampled. Veterinary Certification is dependent on accurate sample identification and correlation.

ii. Tube codes should be entered on the ER16 in the order in which the samples are placed in the aeroboard mould.

iii. ER16 forms should be enclosed within the aeroboard unit in the document compartment.

iv. Aeroboard mould

Order in which samples must be placed in aeroboard mould.

<p>| | | | | |</p>
<table>
<thead>
<tr>
<th></th>
<th></th>
<th></th>
<th></th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>2</td>
<td>3</td>
<td>4</td>
<td>5</td>
</tr>
<tr>
<td>6</td>
<td>7</td>
<td>8</td>
<td>9</td>
<td>10</td>
</tr>
<tr>
<td>11</td>
<td>12</td>
<td>13</td>
<td>14</td>
<td>15</td>
</tr>
<tr>
<td>16</td>
<td>17</td>
<td>18</td>
<td>19</td>
<td>20</td>
</tr>
<tr>
<td>21</td>
<td>22</td>
<td>23</td>
<td>24</td>
<td>25</td>
</tr>
</tbody>
</table>

Needle And Document Compartment
10.5 Description of herd and recording of data on animals sampled

i. It is important to record accurately the information requested on form ER16 in respect of the herd and of each animal from which a blood sample is collected. In particular, it is essential that the PREGNANCY STATUS and abortion history be recorded for each eligible female animal tested.

ii. It is important that the location where the animals are blood sampled is recorded under “address of premises”.

10.6 Forwarding of Samples to Laboratory

i. Samples from more than one herd may be forwarded in the same box provided that the samples from the same herd are not divided among two or more boxes.

ii. It is essential that care be exercised to prevent the box or its aeroboard mould or lid from becoming soiled with blood or other matter.

iii. A separate ER16 form should be completed for each herd represented in a box. Where the number of samples from a herd exceeds 25, a separate ER16 form should be used for each box or part of a box, and each form should be noted “Part of herd of _______ animals” (inserting the total number of samples taken from the herd in question). Separate ER16s are produced on electronic submission.

iv. As for testing for TB, in the case of a herd test all eligible animals on the holding are required to be sampled to obtain the true health status of the epidemiological unit. Accordingly, if there are eligible animals evident to the PVP at the time of the sampling all such animals must be sampled. If this cannot be done the SVI in the RVO must be informed. Brucellosis is extremely contagious and it is critical that if an outbreak occurs all possible infected animals are identified as soon as possible.

v. Place the aeroboard box in the plastic bag provided before placing the whole lot in the cardboard outer box.

vi. The procedure for sealing the box containing the samples is that adhesive security seal labels are provided and one label should be placed over each end of the box.

vii. It is important that the county of the herd tested should be indicated in the space provided on the carton. This greatly simplifies sorting of samples in the laboratory. Filling in the practice name and address in the space provided on the package is also helpful to the laboratory staff.

viii. The blood samples should be forwarded to “The Department of Agriculture, Food and the Marine, Brucellosis Testing Laboratory, Model Farm Road, Cork” by regular post only. Samples incorrectly posted or packed may be destroyed without notification by the postal authorities. The postal authorities also have power to prosecute and recover damages should badly packed blood samples soil other post or personnel.

ix. Samples should as far as possible be posted on the day of collection, but may be posted up to a day after collection. If blood samples must be held over for longer they should be
placed in a refrigerator at 5 degrees centigrade. Blood samples should not be placed in the freezer compartment of a refrigerator.

10.7 Supply of Blood Tubes

i. PVPs will no longer receive routine supplies of blood sampling tubes/kits from the Brucellosis laboratory. Samples and kits submitted for Brucellosis testing will be replaced on a pro-rata basis. PVPs who do not already hold a small float of empty bottles/kits sufficient to perform post-abortion, at risk and a limited number of pre-movement test sampling requirements should contact the Brucellosis laboratory for supply. If a herd test has been listed further supplies as necessary may be obtained on contacting the Brucellosis laboratory. Kits, empty or full, and sample bottles are not provided by ERAD for any purpose other than Brucellosis sampling.

10.8 Disinfection

i. All personnel involved in the testing procedures are required to ensure that proper biosecurity precautions are taken and a thorough cleaning and disinfection of footwear and protective clothing is carried out before leaving the farm premises.

10.9 Removal of Test Materials from Holding

i. It is essential that the residue of all materials employed in the test procedure including syringe covers, used cartons and all other items are gathered and removed at the time of leaving the premises. Safe disposal of used needles is the responsibility of the testing Veterinary Practitioner/VI/Lay Sampler. Under no circumstances should used needles be included in the aeroboard container and sent to the laboratory.

10.10 Private Pre-Movement Testing

i. There is no legal obligation to have animals tested prior to movement but where such testing is being done at the farmer’s request, in order to expedite tests in the above category it is important that the appropriate Private Test label is attached to the outside of the blood sampling kit. These labels should not be used for herd tests.

10.11 General

i. The signature of the testing veterinary practitioner/VI/Lay Sampler on the ER16 must be clearly legible.
Appendix 1 To check handheld re untested animals.

Quport MAHCS Handheld Manual - Page 32

3.5 Viewing Filtered Lists of Animals

MAHCS provides the facility to see a list of animals selected according to a particular filter. By pressing the ‘All…’ button while in Search Mode (make sure keyboard is turned off so you can see the button), you will see a screen like the following:

List Filter

The type of list displayed is selected at the top. Tap on each option to change the list of animals displayed. The list options are:

- Completed – Shows the animals completed for the current day.
- Incomplete – Shows animals not complete for the current day.
- Seen So Far – Shows all the animals found and edited so far. (This is really only of use in BRU-only tests where this test will show all animals that have been seen – even those that were not blood tested)
- Ignored – Lists all ignored animals
- All – Lists all animals (except ignored animals) regardless of what their test status is
- + – Shows all reactor animals
- 0 – Shows all Inconclusive animals

****************************************************************************************************************************


On the Tag screen Press the F4 button to give the following drop down menu. You can use the arrow keys or type T to give the running Totals of the state of play of the test or R to give untested both for Days 1& 2.

group include G group include or exclude e.g. to mark animals for say export certificates or check cards
print P There are a number of print options. Plain paper/ certs etc
wipe W wipes TB and blood bottle readings e.g. for re-test situation.
report R full details of Testing position which is very useful to ensure no readings are missing and other details available from other options displayed e.g. not in group may be used to show card missing.
total T running totals & batches** (7secs.) quick total & batch count of bloods & TB readings.
Dear Sir/Madam

**Bovine Animal identification and associated matters**

The following points cover areas where you have a direct involvement with bovine identification and also areas where you interact with your clients in this and other regards. A number of problems have arisen over the past year which have drawn attention to this area. As you are aware correct bovine identification is very important from the farmers perspective ensuring eligibility for various EU premia. It is also essential for disease control purposes and for maintenance of live cattle exports and beef markets. It is to be expected that various EU mission visits will concentrate more and more over the coming years on identification issues and our compliance with EU regulations in that regard. Your assistance in ensuring that the integrity of bovine identification is secure, in so far as you can, would be most appreciated.

**Correct animal identification and use of brass tags:**

For reasons of traceability and to promote consumer confidence in the wake of the BSE crisis, EU Regulations now require that all bovine animals born since 1.1.1996 are identified by means of two official plastic tags, inserted one in each ear. Up to the end of 1999 calves had to be identified by the keeper within 30 days of birth and registered within 7 days of identification. This interval was shortened to 20 days from 1.1.2000. Tags which become illegible or are lost require replacement by the keeper as soon as is practicable.

When notifying herdowners of your intention to carry out a test it is imperative, therefore, that you remind them that all animals 20 days of age and over must be identified **before the test commences.**

It is mandatory that the herdowner present all animals in the herd at the time of a herd test. However, home-bred animals under 6 weeks of age do not require testing for results to be valid. All animals should be entered on the test report giving the total number for animals presented without identification.

Where **<10%** or 10 animals maximum, over 6 weeks of age, in the herd are not identified then the test should be completed and the unidentified animals provided with temporary identification by the use of a brass tag. ID cards or passports should not be returned to the herdowner until all eligible animals are properly tagged.

Where **>10%** or 10 animals maximum, over 6 weeks of age, in the herd are not identified then a part herd test should be carried out on identified animals only and the remainder of the herd test completed following the identification of those animals in the herd over 20 days of age. The second visit fee involved will be borne by the farmer regardless of who was due to pay the original testing fee.
On the initial day of test you should remind the herdowner of his obligation to tag animals and that he should immediately order tags for this purpose if necessary.

Where animals over 6 weeks of age remain without plastic tags on the day of reading, then the test report, partial or complete, and the passports/identity cards for the herd, must be forwarded to the RVO and attention drawn to the presence of incorrectly identified animal(s). The RVO will arrange to serve a restriction notice for the animals(s) involved and/or the herd as is required by law and take whatever follow-up action is appropriate.

If you have identified, for test purposes, an unidentified animal by means of a brass tag then, of course, no passport (identity card) may subsequently have test details certified by you until such time as you have satisfied yourself as to the correlation between the animal brass-tagged and the plastic tag number on the passport. This ordinarily will require re-visiting the herd to check the animal(s) involved. The herdowner/keeper will be responsible for any costs involved.

Only those animals born prior to the introduction of the new plastic tag identification system may be permanently identified by means of a brass tag. It is the intention, therefore, that brass tags will, within a reasonable timeframe, cease to issue or be available except for the purpose of essential retagging of animals legally brass-tagged.

Passports:

Each animal born in Ireland, after 1.1.98 should have an official pre-printed passport. Obviously the description of an animal on its passport must match the animal. In addition the animal’s keeper is required to sign the passport either on the front if the animal was born into the herd or on the back if purchased. Thus, when you are checking passports at time of test be aware that the herdowner’s/keeper’s signature should ordinarily be on each passport. It is the intention that each animal’s passport will accompany it throughout its life to facilitate completion of a full traceback for consumer protection, fraud prevention, disease control etc. Great care must be taken therefore, to ensure that passports are not lost or mislaid while in your possession.

Farm inspections under the Identification Regulations:

The EU regulations mentioned above also require the competent authority to conduct inspections of a minimum of 5% of holdings, to monitor compliance with the regulations. Operational details for these inspections are currently being finalised. In some instances the RVO may arrange inspections to coincide with TB or other tests being conducted on a herd. Registration records, passport details and entries on the herd register will be checked during these inspections in addition to individual animal tagging. Maintenance of national eligibility for export and other outlets for cattle and meat products and for various EU premia is dependent on compliance with all identification regulations. Therefore, you are advised as part of the general service to your clients to draw their attention to any deficiencies in this regard, that you may observe while on their holdings.
Subject: Bovine Identification and the use of Brass Tags

Purpose

To standardise the manner in which RVOs correlate brass tags used to temporarily identify bovine animals at the time of testing with the animals’ proper plastic tag identification.

Policy

Circular ER6A/2000 that issues annually with the ER4 instructions to all testing PVPs and WTVIs sets out the procedures relating to the treatment of bovines that have been temporarily identified with brass tags. The basis of the veterinary certification contained in that Circular has not changed viz “if the testing vet has identified, for test purposes, an unidentified animal by means of a brass tag then, of course, no passport (identity card) may subsequently have test details certified by the testing vet until such time as they have satisfied themselves as to the correlation between the animal brass-tagged and the plastic tag number on the passport. This ordinarily will require re-visiting the herd to check the animal(s) involved. The herdowner/keeper will be responsible for any costs involved”.

Legislation


Procedures

The AHCS discrepancy report highlights tests where brass tags have been used to temporarily identify animals for the purpose of the test. The ER96 (copy attached) has been revised and is now divided into two sections as follows:

Section A: signed declaration by the keeper regarding the correlation of temporary brass tags with the permanent identity contained on plastic tags.

Section B: signed declaration by the testing veterinary surgeon that the correlation is correct and that he/she can certify the TB and/or Br test.

If the keeper submits the ER96 and Section A only is completed then the discrepancy flag on the AHCS can be lifted by using the facility on the AHCS Edit Animal screen to record the keeper declaration for the temporary tagnumber. Where relevant, the discrepancy flag should also be removed from the permanent tagnumber (in accordance with Council Regulation 1760/2000 the keeper is the person legally responsible for identifying the animal).
If the testing Practitioner has also completed Section B, then the test details may also be updated on AHCS using the Correlate Tags screen and the animal’s passport, if in the RVO, can be returned to the PVP for updating the test details.

The paragraph in the discrepancy letter that refers to animals temporarily tagged has been modified to take account of the changes made to the ER96. A copy of the ER96 should always accompany the discrepancy letter.

The revised ER96 form is now available on the Ezone and all copies of the old version should be withdrawn and destroyed immediately. Supplies of the revised version should also issue to PVPs so that they have copies available to give to the keeper at the time of the test.

The following are the procedures for dealing with animals temporarily tagged at test or indeed animals withheld from test:

1. Details of un-correlated temporary tags (i.e. no ER96 submitted) can be identified by running discrepancy reports for the herd in question.

2. All surplus passports should be forwarded to RVO: this includes passports for animals temporarily tagged at test which are not the subject of an ER96 counter-signed by the testing VS and any animals not presented for test.

3. VS should not retain passports for animals not tested or not presented for test.

4. Passports should be held in the RVO until Section B on the ER96 has been completed by the testing VS.

5. For unregistered animals temporarily tagged at test that will be the subject of a late registration, check that all temporary tags used in the herd have been correlated and that the correlation has been certified by the testing VS, prior to authorizing the issue of a passport.

Key Words: Brass tags, PVPs, Keepers and ER 96

Authorised by: ERAD Management Committee

Date: November 2008
Appendix 3     NOTES ON COMPLETION OF PRE PRINTED ER15B

1. In column headed ‘Breed’ the following entries only should be used:

<table>
<thead>
<tr>
<th>Breed</th>
<th>Code</th>
</tr>
</thead>
<tbody>
<tr>
<td>Angus</td>
<td>AA</td>
</tr>
<tr>
<td>Angler</td>
<td>AN</td>
</tr>
<tr>
<td>Aubrac</td>
<td>AU</td>
</tr>
<tr>
<td>Ayshire</td>
<td>AY</td>
</tr>
<tr>
<td>Belgian Blue</td>
<td>BB</td>
</tr>
<tr>
<td>Bison</td>
<td>BI</td>
</tr>
<tr>
<td>Blone D’Aquitane</td>
<td>BA</td>
</tr>
<tr>
<td>Brown Swiss</td>
<td>BS</td>
</tr>
<tr>
<td>Chianina</td>
<td>CI</td>
</tr>
<tr>
<td>Charolais</td>
<td>CH</td>
</tr>
<tr>
<td>Chianina</td>
<td>CH</td>
</tr>
<tr>
<td>Chianina</td>
<td>CH</td>
</tr>
<tr>
<td>Chianina</td>
<td>CH</td>
</tr>
</tbody>
</table>

2. All animals should be presented with two plastic ear tags for identification purposes. In the case of plastic tags containing an alpha-numeric identifier, the tag number should be written in the format: Letters: numbers: check digit e.g. BEA 19731-4 or Letters: numbers: check letter e.g. BCDF 0025Y. The tag number of plastic tags containing an all-numeric identifier should be written in the order in which the numbers appear on the tag. All zeros included in the number must be recorded. Normally the full space allocated should be used for recording an animal’s tag number.

3. Tag/Pass - The absence of identification documentation (Passport/ID Card) should be indicated by:

(i) NC No Card       ---- Or (ii) FC Full Card ------ Or (iii) WC Wrong Card

Where a temporary tag was used to identify the animal TT should be inserted.

4. Age/DOB should be recorded in the following order – year/month or actual age.

<table>
<thead>
<tr>
<th>Age/DOB</th>
<th>Format</th>
</tr>
</thead>
<tbody>
<tr>
<td>1/0</td>
<td>1 year old</td>
</tr>
<tr>
<td>1/6</td>
<td>one year and six months</td>
</tr>
<tr>
<td>0/10</td>
<td>ten months</td>
</tr>
</tbody>
</table>

5. The following codes should be used for recording the sex of the animal:

<table>
<thead>
<tr>
<th>Sex</th>
<th>Code</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cow</td>
<td>C</td>
</tr>
<tr>
<td>Heifer</td>
<td>H</td>
</tr>
</tbody>
</table>

If an animal is pregnant state the length of pregnancy (e.g. C4 = cow pregnant 4 months).

6. The column headed ‘Clinical Remarks’ is intended for recovering conditions relevant to both TB and Brucellosis testing. In the case of TB the following clinical conditions should be recorded if present:

<table>
<thead>
<tr>
<th>Condition</th>
<th>Code</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cough</td>
<td>CO</td>
</tr>
<tr>
<td>Snoring</td>
<td>SN</td>
</tr>
<tr>
<td>Skin TB</td>
<td>ST</td>
</tr>
</tbody>
</table>

In the case of Brucellosis blood testing any abortion history (i.e. date of abortion) should be entered.

7. In the column headed ‘Reaction’ the following are the appropriate entries.

(i) Diffuse Oedema D.O.    (ii) Extensive Oedema E.O.     (iii) Circumscribed C.

8. In the column headed ‘Result’ you should indicate the result of both increases.

(i) Positive +  Or (ii) Doubtful 0  Or (iii) Negative –

The column headed ‘BR Tested’ must be completed where a prepared list of the animals to be tested is provided by the D.V.O. If an animal has been blood tested the letter ‘Y’ should be inserted in this column to indicate that the animal was blood tested.
**Appendix 4 Cattle Passport**

### Passport/Cattle Identity Card

<table>
<thead>
<tr>
<th>Date of Birth</th>
<th>Tag No.</th>
</tr>
</thead>
<tbody>
<tr>
<td>26 SEP 2010</td>
<td>26/09/2010</td>
</tr>
<tr>
<td>Breed</td>
<td>Sex</td>
</tr>
<tr>
<td>FR</td>
<td>MALE</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>LD. Code of Dam</th>
<th>Date of Issue of Card</th>
</tr>
</thead>
<tbody>
<tr>
<td>IE1234567800059</td>
<td>05/10/2010</td>
</tr>
</tbody>
</table>

**Name & Address of Keeper of Herd/Origin**

JOE FARMER  
GLENROE  
SOMewhere  
CO WICKLOW

**Signature of Keeper**

### Special Beef Premium - Bull Premium Not Claimed

**Certificate of Tuberculosis Testing**

I certify that this animal passed the test indicated below and that no animal failed the test.

<table>
<thead>
<tr>
<th>Date of Test</th>
<th>Herd No.</th>
<th>Signature of Veterinary Surgeon</th>
</tr>
</thead>
<tbody>
<tr>
<td>1.</td>
<td></td>
<td></td>
</tr>
<tr>
<td>2.</td>
<td></td>
<td></td>
</tr>
<tr>
<td>3.</td>
<td></td>
<td></td>
</tr>
<tr>
<td>4.</td>
<td></td>
<td></td>
</tr>
<tr>
<td>5.</td>
<td></td>
<td></td>
</tr>
<tr>
<td>6.</td>
<td></td>
<td></td>
</tr>
<tr>
<td>7.</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

**Certificate of Brucellosis Testing**

I certify that this animal passed the test indicated below and that no animal failed the test.

<table>
<thead>
<tr>
<th>Date of Test</th>
<th>Herd No.</th>
<th>Signature of Certifier</th>
</tr>
</thead>
<tbody>
<tr>
<td>1.</td>
<td></td>
<td></td>
</tr>
<tr>
<td>2.</td>
<td></td>
<td></td>
</tr>
<tr>
<td>3.</td>
<td></td>
<td></td>
</tr>
<tr>
<td>4.</td>
<td></td>
<td></td>
</tr>
<tr>
<td>5.</td>
<td></td>
<td></td>
</tr>
<tr>
<td>6.</td>
<td></td>
<td></td>
</tr>
<tr>
<td>7.</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

---

Stamp test date in any of these 3 columns.